

# TENNESSEE BUREAU OF INVESTIGATION

## Forensic Services Division

### Toxicology Quality Assurance and Procedures Manual

#### 8.7 Benzoyllecgonine Procedure



#### 8.7 BENZOYLECGONINE PROCEDURE

##### 8.7.1 Purpose

To qualitatively and/or quantitatively identify benzoyllecgonine (BE) in submitted evidence using solid phase extraction followed by instrumental analysis with gas chromatography/mass spectrometry (GC/MS).

##### 8.7.2 Specimen Requirements

Samples for analysis will be presumptively positive for cocaine metabolite. Acceptable samples for this analysis include blood and urine. For additional samples see Alternative Matrices (section 6.6).

##### 8.7.3 Apparatus and Equipment

Disposable 10 mL culture tubes  
Volumetric pipettes and disposable tips  
Assorted volumetric glassware  
Disposable transfer pipettes  
Sample mixer  
Centrifuge  
Evaporation station  
Solid phase columns (Bond Elute SCX)  
Solid phase extraction apparatus  
11 mm autosampler vials, inserts, and caps  
11 mm crimper  
GC/MS, ChemStation software, compatible computer, and printer

##### 8.7.4 Reagents and Standards

Negative blood, urine, or other matrix as needed  
Benzoyllecgonine certified reference standard  
Benzoyllecgonine-D<sub>3</sub> certified reference standard (internal standard)  
Water (H<sub>2</sub>O)  
5% Tri-chloroacetic acid (TCA)  
Methanol (CH<sub>3</sub>OH)  
12 mM Hydrochloric acid (HCl)  
12 mM Phosphoric acid (H<sub>3</sub>PO<sub>4</sub>)  
70 mM Glacial acetic acid (GAA) (CH<sub>3</sub>COOH)  
Isopropanol (2-propanol) ((CH<sub>3</sub>)<sub>2</sub>CHOH)  
Acetonitrile (ACN) (CH<sub>3</sub>CN)  
Ammonium hydroxide (NH<sub>4</sub>OH)  
Elution solution (2% triethylamine, 2% NH<sub>4</sub>OH, 20% isopropanol, 76% methylene chloride)



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## Forensic Services Division

### Toxicology Quality Assurance and Procedures Manual

#### 8.7 Benzoylcegonine Procedure

BSTFA + TMCS, 99:1 (Sylon BFT) (N,O-bis(trimethylsilyl)trifluoroacetamide + trimethylchlorosilane)

#### 8.7.5 Standard and Reagent Preparation

The following are examples of how to prepare the standards and reagents used in this procedure.

##### 8.7.5.1 Standards

###### **BE Stock Reference Standard Solution [10,000 ng/mL] (A & B)**

Pipette 500  $\mu$ L of benzoylcegonine [1 mg/mL] certified reference standard and dilute to 50 mL with H<sub>2</sub>O.

###### **BE Stock Reference Standard Solution [1,000 ng/mL] (A)**

Pipette 10 mL of 10,000 ng/mL BE stock reference standard solution (A) and dilute to 100 mL with H<sub>2</sub>O.

###### **BE-D<sub>3</sub> Reference Standard Solution [10,000 ng/mL] (Internal Standard)**

Pipette 500  $\mu$ L of benzoylcegonine [1 mg/mL] certified reference standard and dilute to 50 mL with H<sub>2</sub>O.

###### **BE Working Reference Calibrator/Control Standard Solutions**

To make the working reference calibrator/control standard solutions, add the following amounts to a final volume of 1.5 mL with blood or urine.

CONCENTRATION	AMOUNT USED	STD SOLUTION
50 ng/mL	75 $\mu$ L	1,000 ng/mL
100 ng/mL	150 $\mu$ L	1,000 ng/mL
250 ng/mL	37.5 $\mu$ L	10,000 ng/mL
500 ng/mL	75 $\mu$ L	10,000 ng/mL
1000 ng/mL	150 $\mu$ L	10,000 ng/mL
2500 ng/mL	375 $\mu$ L	10,000 ng/mL
5000 ng/mL	750 $\mu$ L	10,000 ng/mL

##### 8.7.5.2 Prepared Reagents

###### **BE Elution Solution**

Combine 76 mL of methylene chloride, 20 mL isopropanol, 2 mL triethylamine, and 2 mL NH<sub>4</sub>OH.

Note: This solution will expire 24 hours after preparation.



# TENNESSEE BUREAU OF INVESTIGATION

## Forensic Services Division

### Toxicology Quality Assurance and Procedures Manual

#### 8.7 Benzoylcegonine Procedure

##### **5% Tri-chloroacetic acid (TCA)**

Dissolve 25 g of TCA and dilute to 500 mL with H<sub>2</sub>O (also available in a pre-made 5% solution).

##### **12 mM HCl**

Add 983.6 µL HCl to H<sub>2</sub>O and dilute to 1000 mL.

##### **12 mM H<sub>3</sub>PO<sub>4</sub>**

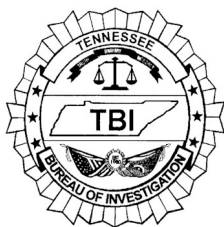
Add 807.5 µL H<sub>3</sub>PO<sub>4</sub> to H<sub>2</sub>O and dilute to 1000 mL.

##### **70 mM GAA**

Add 4.02 mL GAA to H<sub>2</sub>O and dilute to 1000 mL.

#### **8.7.6 Procedure**

1. Allow all reference standards and case samples to equilibrate to room temperature before beginning procedure.
2. Label, check, and load/unload all samples in accordance with the "Sample Pipetting Check List" (see Appendix section).
3. Prepare working reference calibrator and/or control standards from stock reference standards. See example above.
4. Pipette 1.5 mL of corresponding case sample, calibrator, positive control, or negative control into the appropriately labeled 10 mL culture tube.  
Note: Smaller sample volumes may be analyzed on a case-by-case basis.
5. Pipette 37.5 µL of internal standard into each sample to make a final concentration of 250 ng/mL.
6. Add 4 mL of 5% TCA, mix for approximately 90 seconds, and centrifuge until separated (approximately 20 minutes).
7. Transfer the supernatant into another clean, labeled 10 mL culture tube (optional).
8. Activate the column with the following reagents in the following order:
  - 3 mL of methanol
  - 3 mL of 12 mM HCl
9. Add the supernatant to each column slowly (approximately 5-10 psi).
10. Wash the column with the following reagents in order:
  - 1 mL of 12 mM HCl
  - 2 mL of 12 mM H<sub>3</sub>PO<sub>4</sub>
  - 2 mL of 70 mM GAA
  - 3 mL isopropanol
11. Allow the column sorbent to dry completely by allowing flow through the solid phase extraction apparatus for approximately 2 minutes at the highest setting or by changing to "full flow/dry" setting (if available).
12. Switch the solid phase extraction apparatus to the collection tubes.
13. Add 2 mL of elution solution and allow samples to elute slowly (gravity may initially be sufficient, but should be followed by vacuum/pressure).
14. Evaporate the sample to dryness with a dry gas (e.g. nitrogen) in an evaporation station without heat.



# TENNESSEE BUREAU OF INVESTIGATION

## Forensic Services Division

### Toxicology Quality Assurance and Procedures Manual

#### 8.7 Benzoylcegonine Procedure

15. Reconstitute the residue with 50  $\mu$ L of acetonitrile and 50  $\mu$ L of BSTFA+TMCS, mix, transfer to an 11 mm autosampler vial with insert, and seal with cap.
16. Derivatize by heating at approximately 90° for approximately 20 minutes.
17. Analyze and quantitate the samples by GC/MS (selected ion monitoring mode).

#### 8.7.7 Reporting

Results can be reported if the following criteria are met:

**8.7.7.1** Ions 240, 243, 256, 259, 361, and 364 are present in the sample and used calibration standards and control(s).

**8.7.7.2** Ion ratios for the sample must fall within  $\pm 20\%$  of those of a calibrator or control standard of similar concentration.

**8.7.7.3** The retention times of the 361 (benzoylcegonine) and 364 (benzoylcegonine D<sub>3</sub>) ions in the sample are within  $\pm 1\%$  of those of a calibrator or control standard of similar concentration.

**8.7.7.4** Benzoylcegonine concentrations below 50 ng/mL (the lowest calibration point) shall be reported as “not detected”.

**8.7.7.5** Benzoylcegonine concentrations greater than the highest calibration point where the results are necessary for interpretation in the case shall be handled using the following methods:

- Reanalyze using smaller sample amounts (including dilutions).
- Reanalyze using higher standard concentrations.

**8.7.7.6** Benzoylcegonine concentrations greater than the highest calibration point where the results are not necessary for interpretation in the case may be reported as “greater than....” the highest calibration point.

**8.7.7.7** Results shall be reported in ng/mL and truncated to the whole number.

**8.7.7.8** When a definitive conclusion cannot be made, the reason shall be documented on the report (e.g., “insufficient sample for analysis”, “sample unsuitable for analysis”, “results are inconclusive due to sample condition”, etc.).

#### 8.7.8 References

B.K. Logan, D.T. Stafford, I.R. Tebbett, C.M. Moore. “Rapid Screening for 100 Basic Drugs and Metabolites in Urine Using Cation Exchange Solid-Phase Extraction and High-Performance Liquid Chromatography with Diode Array Detection.” Journal of Analytical Toxicology 14 (1990): 154-159.